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Data Article

Guided extracellular matrix formation from fibroblast cells cultured on bio-inspired configurable multiscale substrata



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ABSTRACT

Engineering complex extracellular matrix (ECM) is an important challenge for cell and tissue engineering applications as well as for understanding fundamental cell biology. We developed the methodology for fabrication of precisely controllable multiscale hierarchical structures using capillary force lithography in combination with original wrinkling technique for the generation of well-defined native ECM-like platforms by culturing fibroblast cells on the multiscale substrata [1]. This paper provides information on detailed characteristics of polyethylene glycol-diacrylate multiscale substrata. In addition, a possible model for guided extracellular matrix formation from fibroblast cells cultured on bio-inspired configurable multiscale substrata is proposed.

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1. Specifications table

Subject area	Material science, Physics, Bioengineering
More specific sub- ject area	Extracellular matrix, Scaffold, Cell and tissue engineering
Type of data	Figure and image
How data was acquired	Digital camera and microscope
Data format	Raw and analyzed data
Experimental factors	An equipment for sample preparation Cross-sectional and titled images of samples A proposed biology model
Experimental features	Scanning electron microscope (SEM) samples were placed on a stub for sputter-coating with platinum. Characteristics of samples were analyzed using SEM images. A proposed model for extracellular matrix formation from fibroblast cells was prepared using data.
Data source location	Ulsan National Institute of Science and Technology, Ulsan, Republic of Korea. Chonnam National University, Gwangju, Republic of Korea. Seoul National University, Seoul, Republic of Korea. Ajou University School of Medicine, Suwon, Republic of Korea. Sungkyunkwan University, Suwon, Republic of Korea.
Data accessibility	Data are provided in this paper and related to [1].

2. Value of the data

- Detailed figures for the methodology for fabrication of precisely controllable multiscale hierarchical structures was provided.
- Detailed data on the height and cross-sectional characteristics of the multiscale substrata were provided.
- A possible model for multiscale topographical cues-guided extracellular matrix formation from fibroblast cells was proposed.

3. Data, experimental design, materials and methods

Hierarchically polyethylene glycol-diacrylate (PEG-DA) multiscale patterned substrata were fabricated using our developed method [1] including capillary force lithography in combination with original wrinkling technique (Figs. 1 and 2).

To enhance the adhesion strength between the PEG-DA layer and the UV/O-treated polydimethylsiloxane (PDMS) sheet, we used 3-trimethoxysilylpropyl methacrylate (TMSPMA) as an adhesion promoter. The acrylic functional group of the TMSPMA covalently bonded with the oxidized PDMS and the PEG-DA, enabling a strong adhesion between the different layers (Fig. 3).

Fig. 4 shows the representative scanning electron microscope (SEM) images of a multiscale substrate. The height of micowrinkle pattern is $\sim 30\text{ }\mu\text{m}$ and the height of nanogroove pattern is $\sim 250\text{ nm}$.

Using bio-inspired configurable multiscale substrata, we cultured NIH3T3 fibroblast cells with Dulbecco's Modified Eagle's medium (DMEM) with 10% fetal bovine serum (FBS) and 1% penicillin–

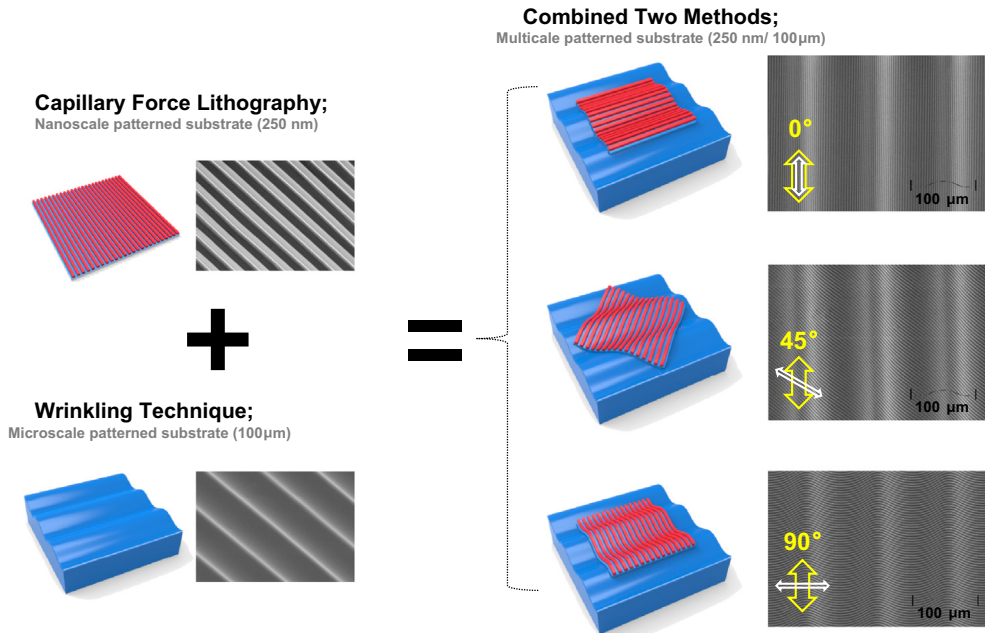


Fig. 1. A schematic for the methodology for fabrication of precisely controllable multiscale hierarchical structures based on the two fabrication methods such as capillary force lithography and wrinkling technique. The yellow arrows indicate the direction of micro scale structures and the white arrows indicate the direction of nanoscale structures in the SEM images.

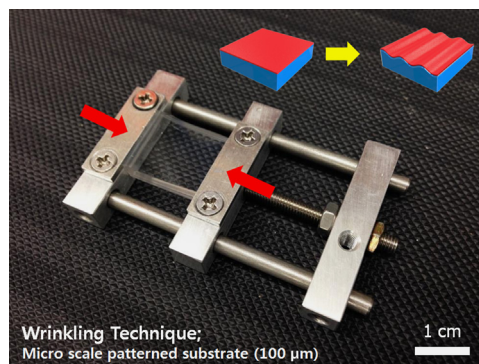


Fig. 2. A photograph image of the custom designed strain apparatus used in this work. The red arrows indicate the strain direction.

streptomycin (Gibco, Milan, Italy) at 37 °C in a 5% CO₂ atmosphere for engineering complex ECMs [1]. It was hypothesized that controlled shape and orientation of fibroblast cells on multiscale topography could affect the production of ECM molecules from them [1–4]. We showed that the multiscale patterned substrata could guide the shape and orientation of NIH3T3 fibroblast cells as well as significantly improve the secretion of fibronectin from them than the micro wrinkle patterned substrata [1]. This suggests that bio-inspired configurable multiscale substrata could guide the ECM formation from fibroblast cells for engineering native-like ECM platforms for advanced tissue engineering (Fig. 5).

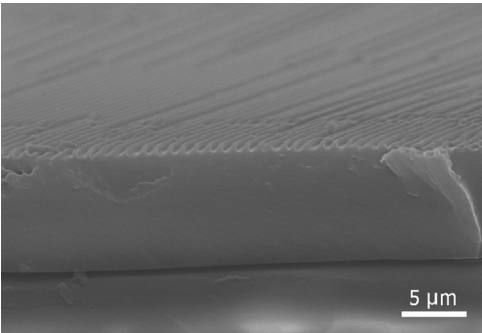


Fig. 3. A representative SEM image of a multiscale substrate composed of nanopatterned PEG-DA upper layer and UV/O treated PDMS lower layer. These two layers are covalently bonded using TMSPMA adhesion promoters.

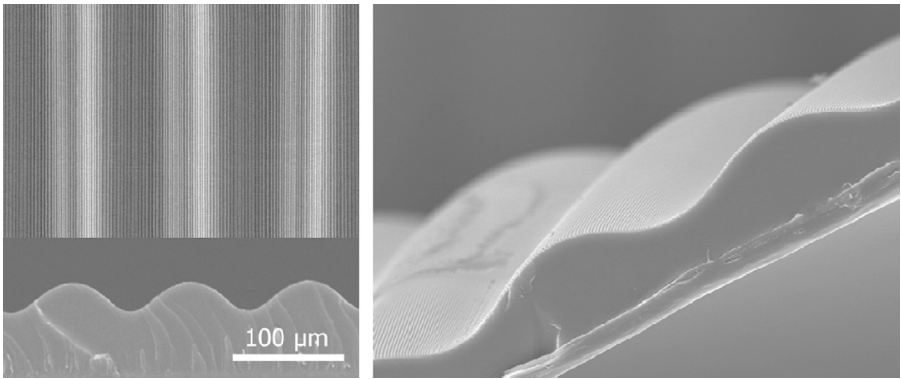


Fig. 4. Top, cross-sectional and titled SEM images of a multiscale substrate. The height of micro wrinkle pattern is $\sim 30\ \mu\text{m}$ and the height of nanogroove pattern is $\sim 250\ \text{nm}$.

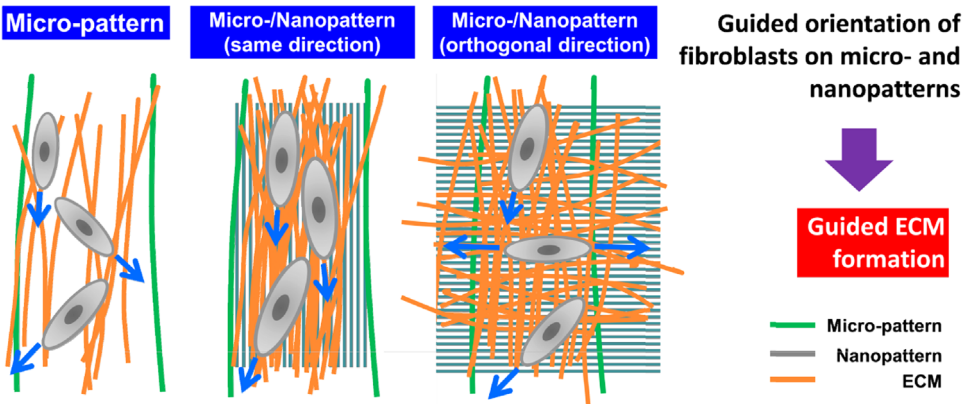


Fig. 5. A possible model for guided extracellular matrix formation from fibroblast cells cultured on bio-inspired configurable multiscale substrata.

Acknowledgments

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References

- [1] W.G. Bae, J. Kim, Y.H. Choung, Y.S. Chung, K.Y. Suh, C. Pang, J.H. Chung, H.E. Jeong, Bio-inspired configurable multiscale extracellular matrix-like structures for functional alignment and guided orientation of cells, *Biomaterials* 69 (2015) 158–164.
- [2] H.N. Kim, A. Jiao, N.S. Hwang, M.S. Kim, D.H. Kang, D.H. Kim, K.Y. Suh, Nanotopography-guided tissue engineering and regenerative medicine, *Adv. Drug Deliv. Rev.* 65 (2013) 536–558.
- [3] J. Kim, H.N. Kim, Y. Lang, A. Pandit, Biologically inspired micro-and nanoengineering systems for functional and complex tissues, *Tissue Eng. Part A* 20 (2014) 2127–2130.
- [4] D.H. Kim, P.P. Provenzano, C.L. Smith, A. Levchenko, Matrix nanotopography as a regulator of cell function, *J. Cell Biol.* 197 (2012) 351–360.